



## METHOD TO OBTAIN MICROPARTICLES

Field of invention

The present invention provides a method of obtaining microparticles by a spray freezing technique. More specifically the present invention relates to a method by which spherical microparticles containing one or more pharmaceutically active substances can be prepared.

Background of the invention

The strategy for pharmaceutical formulation of a given drug depends on different factors. Ultimately, these factors emanate from 1) the therapeutic needs, 2) the physical chemical properties of the drug, and 3) the influence of the biological environment where the formulation will release its contents. Thus, both technical and biopharmaceutical considerations will contribute to a successful therapy.

However, improved drug administration may also be achieved by so called modified release of the drug, which has been discussed extensively in the literature, *e g R L Langer and D L Wise (Eds) "Medical Applications of Controlled Release", vols I, II (1984), CRC Press Inc, Boca Raton.*

Several approaches to achieve different types of modified release are described in the references above. Of special importance to the present invention is modified release achieved by formulating the active substance with a suitable carrier material in the form of microparticles. Such a formulation then contains multiparticulate discrete delivery units, each of which can be coated if necessary with, *e g* a suitable pH sensitive, semipermeable or other polymeric film. Several advantages can be obtained with this type of formulation compared with more conventional delivery means. Thus, the small size of the

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microparticles assures a fast and predictable emptying from the stomach, which is of special importance in the presence of food. Further, the particles will spread over a larger area in the whole GI-tract compared with a conventional monolithic (single-unit) formulation. This will result in a safer therapy when the active substance has local irritating side effects. Controllable plasma levels of absorbed drug can also be obtained. The microparticle formulation will also have a longer residence time in the colon which makes 24 hrs extended release formulations possible. From a technological point of view, microparticles are more suitable for coating and handling since a technical fault during the process may be serious for single unit formulations but less so for micropellets. Also, microparticle formulations are more easily manufactured and prepared in different doses than standard tablet systems.

#### Prior Art

An ideal method for the preparation of microparticles where the drug is homogeneously distributed within a polymeric matrix, should be simple, reproducible, rapid and minimally dependent on the solubility characteristics of the drug. A high product yield and a high degree of retention of the active substance in the final microparticles should also be obtained.

Several different techniques are available for making microparticles (< 1 mm), e g spray-drying, extrusion-spheronization, spray-chilling, emulsion solvent evaporation/extraction and coating of nonpareil spheres, among others. A recent review was presented by Conti et al in STP Pharma Sci 7, 331 (1997) where the technical aspects of coacervation, spray-drying, emulsion solvent extraction, and emulsion solvent evaporation were discussed.

However, all existing techniques suffer from one or more drawbacks. Thus, many drugs are sensitive to heat and therefore will deteriorate which restricts the use of spray-drying or spray-chilling.

In extrusion spheronization and in coating of non-pareils particles it has been difficult to achieve acceptable microparticles in the size range of 50 - 400  $\mu\text{m}$ . Pellets made by these methods contain significant amounts of inert excipients. This may make the pelletization of high-dose drugs by these methods a difficult task.

Finally, in emulsification solvent evaporation, an emulsion has to be made and the drug to be incorporated is preferably lipophilic, which restricts the drugs which can be used. Another drawback is the toxicity of the solvent used, usually methylene chloride, which can remain in the microparticles after drying.

However, despite the many different approaches there has not been disclosed a technique that can produce both smaller microparticles but also particles of more uniform size. It is important to avoid, e g segregation and dose variation during further processing into capsules or tablets. Further, the existing techniques do not incorporate several desirable aspects such as the possibility to produce spherical microparticles of different size ranges that are homogeneous, have a high drug content and sufficient mechanical strength ( to e g withstand coating processes) into one single technique.

A spray-freezing technique has been used for the processing and granulation of ceramic materials to achieve homogeneous distribution of additives within granules to be compacted. For the processing of slurries containing silicon-nitride, sintering additives and a binder, spherical free-flowing granules have been prepared by spray-freezing and subsequent freeze-drying. The homogeneity of the slurry was retained in the granules and thus in the final sintered product (Nyberg et al, Euro-Ceramics II 1, 447 (1993)).

Suspensions of silicon carbide and additives were processed in this way to give granules for compaction (US Patent 4,526,734). The increased homogeneity compared with traditional granulation techniques resulted in better mechanical properties of a whisker reinforced ceramic (EP 0 584 051). The process is also feasible for making homogeneous powder blends for ceramic superconductors (Japanese unexamined patent application no. 59-102433).

Normally pharmaceutical materials are lyophilized by freeze-drying in a bulk process in which the solution/suspension to be freeze-dried is placed in vials or on trays in a freeze-drier, where freezing and subsequent sublimation of the dry solvent take place. The dried product is a powder cake.

The rapid freezing provided by spray-freezing ensures that no concentration gradients exist in the resulting frozen particles and degradation of biological material is prevented. This approach has been used to achieve precise metering and dispensing (M J Akers and D J Schmidt, BioPharm 28, (April 1997)); where the frozen particles were in the form of large lumps of size 1-9 mm. Freezing of droplets in a moving bath of Freon 12 (-20°C), which medium conflicts with environmental demands, has been used to obtain porous, free-flowing, spherical granules with rapid dissolution (US patent 3,932,943); as well as making homogenous granules for tableting with precise dosing (US Patent 3,721,725).

A process for preparing foamed bioabsorbable polymer particles for surgical use was presented in US Patent 5,102,983. Here, however, the porosity was very large, and the pore sizes in the range of 4 - 10  $\mu\text{m}$ , the dry content of the solution being sprayed being 1 - 20 wt%.

US Patent 5,019,400, disclosed the use of a mixture of a biologically active material, a polymer, and a solvent which was sprayed into a non-solvent cooling medium that froze the droplets with subsequent extraction of the solvent in the droplets during heating. The particles were finally dried in a vacuum-drier. The microparticles formed were porous, and contained 0.01 - 50 % of the active substance. The dry content of the solution sprayed was 6 wt%. This process is not entirely satisfactory since it is an advantage to have one single drying step after freezing and also a higher active substance content than 50 wt% in order to make high dose materials.

US Patent 5,405,616 discloses a method of forming droplets by forcing a suspension/solution/emulsion through calibrated jets. The droplets then fall into liquid nitrogen. Due to low shear forces the size of the pellets formed is large; 0.2 - 12 mm, which would then give a less safe dosability than if smaller particles could have been achieved. The smallest particles achieved were 0.8 - 1 mm. Further, to achieve low friability pellets, the drying step after freeze-drying was performed by thawing the pellets before conventional vacuum drying. To achieve these low friability pellets the matrix former is restricted to materials that during thawing will form a gel. The particles obtained contain no more than 33 wt% of the active substance.

To the skilled person particle production utilizing the technique described in US 5,405,616 appears to be quite a slow process and not suitable for large scale industrial pharmaceutical production.

#### Object of the invention

An object of the present invention is to provide a method for the production of microparticles. More specifically, the method is for the production of homogeneous microparticles which does not have the drawbacks of the methods discussed above, e.g. methods that rely on heat or multiple solvents for drug dissolution, but instead puts no restrictions on the drug to be incorporated. A further object is to provide a method for the production of microparticles with controllable amounts of incorporated drug in a high-yield process. Also, the invention provides a method to produce homogeneous microparticles with an incorporated drug that have low friability so that they for instance can withstand coating processes. A further object of the invention is to provide a method to produce microparticles that have easily controllable density and strength. A further object is to obtain microparticles with a high content of active substance.

### Disclosure of the invention

It has now been found that free-flowing, homogeneous microparticles having low friability can be obtained by spray-freezing a suspension, solution or emulsion of a pharmaceutically active substance with subsequent freeze-drying of the frozen microparticles. The microparticles are preferably spherical in shape. The porosity of the microparticles obtained is controlled in the process by the dry content of the suspension, solution or emulsion. Apart from the porosity, the brittleness of the microparticles is controlled by the amount of polymer binder included in the suspension, solution or emulsion. In order to obtain low friability particles the dry content of the suspension or solution or emulsion should be high.

Generally the following conditions are applicable to obtain low friability microparticles according to the method of the invention;

Low friability microparticles, that can for instance withstand coating with a polymeric film, are achieved when the suspension, solution or emulsion has a dry volume content of at least 15 vol%, preferably up to 60 vol %, and a polymer binder content of at least 5 weight %, preferably 10 weight % or more, and more preferably 15 weight % or more (based upon dry content). A high total pharmaceutically active substance content can be obtained by using the present invention, such as up to 95 weight % or preferably 90 weight % (based upon dry content). The median pore size of the microparticles obtained being preferably a maximum of 1.0  $\mu\text{m}$ . Dry content and dry volume content are weight % and volume %, respectively, of dry material in the suspension/solution/emulsion (dry/(dry + liquid)), wherein the dry material is pharmaceutically active substance + polymer.

According to the present invention homogenous low friability microparticles can be obtained when the dry content is from 15 to 60 vol % and the polymer binder content is 5 weight % or more giving dry microparticles with a relative density of 15 to 60 % (a

porosity of 85 down to 40 vol %). [Relative density: weight of freeze-dried material/volume of freeze-dried material/theoretical density of dry material].

The content of the pharmaceutically active substance calculated on the weight of the dried  
5 microparticles may be from 60 to 95 weight %, preferably from 75 to 90 weight %.

The dry content of the liquid medium is defined as the residue after drying at 110°C for  
2 hours, divided by the total amount before drying. The dry content can be expressed either  
as weight percent or, preferably, as volume percent.

The success in obtaining low porous microparticles and thus low friable microparticles  
depends on the volume fraction of dry material and the amount of polymer binder. The dry  
content of a suspension/solution/emulsion should thus preferably be expressed as a volume  
fraction although this cannot always be calculated.

The microparticles may be obtained by spraying a homogeneous suspension, solution or  
emulsion of the active substance(s) through an atomizer into a vessel with a cold medium  
with a temperature well below that of the freezing point of the liquid in the droplets.  
Frozen droplets will then form instantaneously. The structure of the suspension, solution or  
20 emulsion is retained in the droplets providing a homogeneous distribution of the substances  
within the droplets. The frozen liquid is then sublimated by freeze-drying of the frozen  
droplets where the structure of the droplets is retained due to lack of migration of  
substances during drying.

25 The following general steps of the procedure are further exemplified in the Experimental  
Section below :

a) Preparation of a medium for atomizing. The medium is a suspension, a solution or an  
emulsion of the active substance. A suspension may be prepared by dissolving or  
30 dispersing a polymer in a liquid (as defined below), and then adding fine particles of the

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active substance. A further dispersing agent ( typically in an amount of less than 20 % (w/w) of the polymer amount) might also be included to facilitate the dispersion of the active substance. The polymer might then act as a binder between the fine active substance particles in the microparticles and can be either a water soluble or a non-water soluble polymer, according to definitions below.

b) Atomizing of the suspension/solution/emulsion into droplets. The suspension, solution or emulsion is fed by e.g. a peristaltic pump through a nozzle that could be a pneumatic nozzle, an ultrasonic nozzle, a rotary atomizer or a pressurized nozzle. A typical size distribution of spheres produced by this process can range from 1000  $\mu\text{m}$  down to 10  $\mu\text{m}$ .

c) Freezing of the formed droplets: The atomizer is situated above the cold medium in a cylindrical vessel. If the cold medium is a liquified gas the droplets in the spray formed by the nozzle hit the cold boiling gas before hitting the cold medium that is stirred to get a better wetting of the droplets. Instant freezing takes place and the structure of the homogeneous suspension is retained within the frozen microparticles.

d) Sublimation of the frozen liquid within the droplets: The frozen droplets are transferred from the cold medium to a freeze-drier to sublimate the frozen liquid. This step takes place without any shrinkage of the droplets or migration of excipients ( e g polymers) and thus the structure of the suspension/solution/emulsion is retained within the dry microparticles.

The polymer or dispersing agent used for the formulation may be a dry polymer that is partly or fully soluble in the liquid. The polymer or dispersing agent used might also be a dispersion of polymer particles (e g a latex).

The polymer or dispersing agent could be but are not limited to the excipients listed below.

- *cellulose derivatives*, like ethylcellulose, hydroxypropyl methyl cellulose, hydroxyethyl cellulose, hydroxypropyl cellulose, ethyl hydroxyethyl cellulose, carboxymethyl cellulose, cellulose acetate butyrate, cellulose acetate phtalate, methylcellulose, etc

5 - *other polysaccharides*, like alginate; xanthan; carrageenan; scleroglucan; pullulan; dextran; hyaluronic acid; chitin; chitosan; starch; etc

- *other natural polymers*, like proteins (e g albumin, gelatin, etc); natural rubber ; *gum arabic*; etc

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- *synthetic polymers*, like acrylates (e g polymethacrylate, poly(hydroxy ethyl methacrylate), poly(methyl methacrylate), poly(hydroxy ethyl methacrylate - co methyl methacrylate), Carbopol® 934, etc); polyamides (e g polyacrylamide, poly(methylene bisacrylamide), etc); polyanhydrides (e g poly(bis carboxyphenoxy)methane, etc); PEO-  
15 PPO block-co-polymers (e g poloxamers, etc); polyvinyl chloride; polyvinyl pyrrolidone; polyvinyl acetate; polyvinyl alcohol; polyethylene, polyethylene glycols and co-polymers thereof; polyethylene oxides and co-polymers thereof; polypropylene and co-polymers thereof; polystyrene; polyesters (e.g. poly(lactic acid), poly(glycolic acid), poly(caprolactone), etc, and co-polymers thereof, and poly(ortho esters), and co-polymers  
20 thereof); polycarbonate; cellophane; silicones (e.g. poly (dimethylsiloxane), etc); polyurethanes; synthetic rubbers (e.g. styrene butadiene rubber, isopropene rubber, etc); etc

- *surfactants*, e.g. anionic, like sulphated fatty alcohols (e g sodium dodecyl sulphate),  
25 sulphated polyoxyethylated alcohols or sulphated oils, etc; cationic, like quaternary ammonium and pyridinium cationic surfactants, etc; non-ionic, like polysorbates (e.g. Tween), sorbitan esters (e.g. Span), polyoxyethylated linear fatty alcohols (e.g. Brij), polyoxyethylated castor oil (e g Cremophor), polyoxyethylated stearic acid ( e g Myrj), etc.

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The mechanical strength of the microparticles is important for determining whether they will withstand processing with a polymer coating in a fluid bed.

Examining the microparticles with a microscope before and after the fluidization in a fluid bed will give an indication of their mechanical strength.

5 To achieve a relative measurement of mechanical strength the pressure where microparticles started to deform was evaluated. Microparticles within a certain size range (sieve fraction) were placed as a monolayer onto the surface of a probe with a certain area. Different loads (forces) were applied to the layer of microparticles for one minute.

10 Examination of the monolayer of microparticles before and after loading was made in a Scanning Electron Microscope to see at what load the microparticles started to deform. The pressure at which the microparticles started to deform was then calculated.

Pharmaceutically active substances suitable to form microparticles of this invention can be  
15 but are not limited to peptides, proteins, low molecular organic substances, pro-drugs, antigens, hormones.

Thus, a microparticle according to the present invention comprises one (or several) pharmaceutically active substances with one or several additional non-active substances,  
20 which are dispersed within the microsphere.

Uncoated particles can be retrieved as they are easily dissolved when they are immersed into a liquid due to their porous structure.

25 The microparticles obtained can be coated with a polymer to achieve either a time-controlled release, a site-controlled release or a pH-dependent release. Suitable polymers for coating can be, but are not limited to, the same type of polymers as listed above.

The coated microparticles can be put into capsules or incorporated into a tablet compressed  
30 by methods known by those skilled in the art.

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The formulations produced based on the microparticles, coated or uncoated, can be given by different administration routes, such as, but not limited to, the oral, the parenteral, the nasal, the pulmonary, the rectal, the tonsillar, the buccal, the intraocular, the vaginal etc, administration routes. The preferred administrations are by the oral, nasal, pulmonary and rectal routes.

### Working examples

The following examples illustrate different aspects of the invention.

The size distribution of the obtained microparticles was measured by sieving. By mercury porosimetry measurements the bulk-density and pore-size distributions were determined. To determine the median pore size the pressure range for mercury intrusion corresponded to pore sizes between 0.0005  $\mu\text{m}$  and 10  $\mu\text{m}$ .

By subjecting a monolayer of the microparticles to compaction forces their relative strength was measured.

**EXAMPLE 1: Preparation of microparticles with a high loading of dry content that withstand coating in a fluidized bed**

A suspension containing talc powder was made according to the composition below;

Talc powder (1-2 $\mu\text{m}$ )	90 g
Talc powder (5 - 80 $\mu\text{m}$ )	210 g
HPMC, 6 cps	80 g
Tween 80 (polysorbate 80)	6 g
Purified water	750 g

Weight percent of dry content in suspension: 34 (19.2 vol%)

First, polysorbate 80 was mixed with the water. The HPMC was then added and dissolved during stirring with subsequent addition of the substance. The suspension was then  
5 deagglomerated by high-shear mixing. The deagglomerated suspension was fed through a pneumatic nozzle with a diameter of 1.0 mm at a speed of about 15 ml/min. The pressure of the atomizer was 1 bar. The spray formed first hit the cold gas above the liquid in a vessel filled with liquid nitrogen that was stirred to get a better wetting and instantaneous freezing of the droplets. The frozen droplets have a higher density than liquid nitrogen  
10 which make them sink to the bottom of the vessel. The frozen droplets/microparticles were then placed in a conventional freeze-drier with a shelf-temperature of -20°C. The primary drying was performed stepwise at -20°C to 0°C at 0.1 mbar. The dry microparticles were free-flowing and spherical. Scanning Electron Microscopy showed a homogeneous distribution of the talc powder with pores (0.1 - 2 µm) in between. The bulk density,  
15 median pore size and mechanical strength was measured and the results are shown in table 2.

Compaction measurements showed that the microparticles obtained had a low friability (high mechanical strength).

20 Fluidization of the microparticles in Example 1 in a fluidized bed showed by microscopy that the microparticles did not break down. These microparticles started to deform at a pressure of 94 kPa (sieve fraction: 450-630 µm). Final coating with a polymer in a fluidized bed proved that the microparticles could be successfully coated.

#### 25 EXAMPLE 2: Coating of microparticles with a polymeric film

The microparticles from Example 1 were easily handled without falling apart and tough enough to be successfully coated. A fraction of 20 g of the microspheres, 150 - 300 µm in

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**Characterization of pellets obtained in example 1.**

Table 1. Size distribution. Sieving (weight fraction%)

Fraction	Example 1
< 100 $\mu\text{m}$	1
100 - 150 $\mu\text{m}$	2
150 - 300 $\mu\text{m}$	22
300 - 450 $\mu\text{m}$	32
450 - 630 $\mu\text{m}$	26
630 - 800 $\mu\text{m}$	12
800 - 1000 $\mu\text{m}$	4

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Table 2. Characterization of microparticles

Example no.	Dry content (vol%)	Binder (wt%) based on dry content	Mercury porosity measurements		Mechanical strength	
			Bulk density (g/cm <sup>3</sup> )	Pore median size ( $\mu\text{m}$ ) (measured range: 0.0005- 10 $\mu\text{m}$ )	Kpa	Fraction
1	19.2	21	0.47	0.8	94	450 - 630 $\mu\text{m}$

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